

Antibacterial, *In Vitro* Analgesic, and Antioxidant Activities of Crude Extract of *Habenaria plantaginea* Lindl.

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Abstract

The rising problem of antibiotic resistance and oxidative stress-related disorders have increased the need for natural sources of therapeutic agents. *Habenaria plantaginea* Lindl., a medicinally important orchid, is known to contain bioactive phytochemicals with potential pharmacological effects. This study evaluated the antibacterial, in-vitro COX-2 inhibitory, and antioxidant activities of the crude root extract of *H. plantaginea*. Antibacterial activity was assessed against five clinically significant bacterial strains (*Escherichia coli*, *Klebsiella pneumoniae*, *Staphylococcus epidermidis*, *Staphylococcus aureus*, and *Bacillus subtilis*) using the disc diffusion method, with streptomycin as a standard. The crude extract exhibited broad-spectrum antibacterial activity, showing the highest inhibition against *B. subtilis* (26.89 ± 1.00 mm) and *E. coli* (24.50 ± 1.29 mm). In the COX-2 inhibitory assay, the extract demonstrated moderate, concentration-dependent inhibition with a maximum of $42.36 \pm 1.18\%$ at $250 \mu\text{g/mL}$ ($\text{IC}_{50} = 190.2 \pm 2.5 \mu\text{g/mL}$), whereas the standard drug celecoxib achieved $94.21 \pm 1.10\%$ inhibition. Similarly, the extract showed moderate antioxidant activity in the H_2O_2 scavenging assay, with a maximum inhibition of $46.38 \pm 1.30\%$ at $250 \mu\text{g/mL}$ ($\text{IC}_{50} = 205.4 \pm 3.1 \mu\text{g/mL}$), compared to $91.45 \pm 1.12\%$ for ascorbic acid. These results suggest that the crude extract of *H. plantaginea* possesses significant antibacterial, anti-inflammatory, and antioxidant potential, supporting its traditional use and highlighting its promise as a source of bioactive compounds for pharmaceutical applications.

KEYWORDS

Antibacterial activity, COX-2, H_2O_2 , *Habenaria plantaginea* Lindl.

1.0 INTRODUCTION

The growing burden of infectious diseases, pain-related disorders, and oxidative stress-associated complications continues to pose a serious threat to global public health [1-2]. Bacterial infections remain a major cause of morbidity, while pain and inflammation significantly reduce quality of life. In parallel, oxidative stress resulting

from an imbalance between reactive oxygen species (ROS) and antioxidant defense mechanisms plays a key role in the pathogenesis of numerous chronic diseases, including inflammation, cancer, diabetes, and neurodegenerative disorders. These challenges highlight the urgent need for safer and more effective therapeutic agents [1, 3].

Medicinal plants have long served as an important source of bioactive compounds with antimicrobial, analgesic, and antioxidant properties. Plants synthesize a diverse range of secondary metabolites such as alkaloids, flavonoids, phenolics, terpenoids, glycosides, and tannins, many of which exhibit significant pharmacological activities [4-5]. Due to their structural diversity and multi-target effects, plant-derived compounds continue to attract scientific interest as potential alternatives or complements to synthetic drugs. *Habenaria plantaginea* Lindl., a member of the family Orchidaceae, is a medicinally important terrestrial orchid traditionally used in various systems of folk medicine. The plant has been employed for the management of pain, inflammation, wounds, infections, and general weakness [6-7]. Despite its traditional relevance, scientific validation of its pharmacological properties remains limited, particularly regarding its antibacterial, analgesic, and antioxidant potential. Previous phytochemical studies on species of the genus *Habenaria* have reported the presence of bioactive constituents such as phenolic compounds, flavonoids, sterols, and glycosides, which are known to contribute to antimicrobial, free-radical scavenging, and analgesic effects. However, systematic evaluation of the biological activities of crude extracts of *H. plantaginea* is still scarce [8].

Therefore, the present study was designed to investigate the antibacterial sensitivity, in-vitro analgesic, and antioxidant activities of crude extracts of *Habenaria plantaginea* Lindl. The findings of this work aim to provide scientific evidence supporting the traditional use of the plant and to explore its potential as a natural source of therapeutically valuable bioactive compounds.

2.0 MATERIALS AND METHODS

Fresh roots of *Habenaria plantaginea* Lindl. were collected from District Dir, Khyber Pakhtunkhwa, Pakistan. The plant material was authenticated by a qualified botanist. The collected roots were thoroughly washed with distilled water to remove adhering soil and impurities, shade-dried at room temperature, and then ground into a fine powder using a mechanical grinder [9].

2.1 Extraction

Approximately 250 grams of powdered root material was subjected to cold maceration using analytical grade methanol for 72 hours with periodic stirring. The extract was filtered and concentrated using a

rotary evaporator under reduced pressure at a controlled temperature.

2.2 Antibacterial Assay

The disc diffusion method was employed to assess the antibacterial activity. Bacterial cultures (*E. coli*, *K. pneumoniae*, *S. epidermidis*, *S. aureus* and *B. subtilis*) were adjusted to 0.5 McFarland standard. Sterile discs impregnated with individual extracts were placed on pre-inoculated agar plates. Streptomycin served as a positive control. Plates were incubated at 37°C for 24 hours, and the inhibition zones were measured (mm).

2.3 COX-2 assay

The COX-2 inhibitory assay was carried out to evaluate the anti-nociceptive activity by using the previously reported procedure [10]. Prepared the enzyme solution having a concentration of 300 U/ml. Celecoxib was used as a positive control in the current assay. Activation of enzyme solution (10 µl) was carried out by using the ice for 5 min with 50 µl cofactor solution containing (glutathione 0.9 mM), TMPD (*N,N,N,N*-tetramethyl-p-phenylenediaminedihydrochloride) (0.24 mM) and hematin (1 mM) in Tris buffer (0.1 M) with pH 8. Kept the Enzyme solution (60 µl) and test samples (20 µl) with various concentrations for five minutes at room temperature. The absorbance of the samples was measured at 570 nm after the samples have been incubated for 5 minutes. The reaction was initiated by adding arachidonic acid (30 mM, 20 µl). The percent inhibition was analyzed from absorbance value per unit time.

2.4 H₂O₂ Assay

As per our previously reported method, the H₂O₂ scavenging activity of tested sample was elucidated. A solution of H₂O₂ (2 mM) was prepared in phosphate buffer (50 mM, pH 7.4). In a 0.3 mL of phosphate buffer solution (50 mM), 0.1 mL of test sample was added, which was followed by addition of 0.6 mL of H₂O₂ and then vortexed. The absorbance of solution was measured at 230 nm after a period of 10 min (UV-3000 O.R.I. Germany), in comparison to blank [11-12].

2.5 Statistical Analysis

All tests were performed in triplicate and values were represented as mean ± standard deviation. Results were interpreted based on concentration-dependent activity patterns.

3.0 RESULTS

3.1 Antibacterial activity

The antibacterial activity of the crude extract of *Habenaria plantaginea* Lindl. was evaluated against selected Gram-positive and Gram-negative bacterial strains using streptomycin as a standard antibiotic (Table 1). The crude extract exhibited notable antibacterial activity against all tested microorganisms, though the degree of inhibition varied among the strains. Against *Escherichia coli*, the crude extract produced an inhibition zone of 24.50 ± 1.29 mm, which was slightly lower than that of streptomycin (27.76 ± 1.05 mm). In the case of *Klebsiella pneumoniae*, the extract showed appreciable activity with an inhibition zone of 23.56 ± 1.50 mm, compared to 31.23 ± 1.49 mm exhibited by the standard drug. For Gram-positive bacteria,

the crude extract demonstrated moderate to strong antibacterial effects. The inhibition zone recorded against *Staphylococcus epidermidis* was 16.34 ± 1.34 mm, whereas streptomycin showed significantly higher activity (35.11 ± 1.23 mm). Against *Staphylococcus aureus*, the crude extract exhibited an inhibition zone of 20.65 ± 1.65 mm, compared to 29.60 ± 1.20 mm for streptomycin. The strongest antibacterial response of the crude extract was observed against *Bacillus subtilis*, with an inhibition zone of 26.89 ± 1.00 mm, while the standard antibiotic produced 34.39 ± 1.77 mm. Overall, the results indicate that the crude extract of *Habenaria plantaginea* possesses broad-spectrum antibacterial activity, with comparatively stronger effects against *Bacillus subtilis* and *Escherichia coli*. The antibacterial activity results are summarized in Table 1.

Table 1. Antibacterial activity of crude extracts of *Habenaria plantaginea* Lindl.

Bacterial strain	Crude	Streptomycin
<i>E. coli</i>	24.50±1.29	27.76±1.05
<i>K. pneumonia</i>	23.56±1.50	31.23±1.49
<i>S. epidermis</i>	16.34±1.34	35.11±1.23
<i>S. aureus</i>	20.65±1.65	29.60±1.20
<i>B. subtilis</i>	26.89±1.00	34.39±1.77

3.2 COX-2 Assay Result

The in-vitro COX-2 inhibitory activity of the crude extract was evaluated and compared with celecoxib. Celecoxib showed strong, concentration-dependent inhibition with a maximum of $94.21 \pm 1.10\%$ at $250 \mu\text{g/mL}$ and an IC_{50} value of $11.8 \pm 0.7 \mu\text{g/mL}$. The crude extract exhibited moderate inhibition, producing $42.36 \pm 1.18\%$ inhibition at the highest concentration tested. A

gradual decrease in inhibitory activity was observed with decreasing concentrations. The IC_{50} value of the crude extract was calculated as $190.2 \pm 2.5 \mu\text{g/mL}$. The inhibitory effects of the crude extract were statistically significant ($***p < 0.001$) compared to the control as shown in table 2.

Table 5: In-vitro Cox-2 inhibitory activity

Sample	Conc ($\mu\text{g/mL}$)	% Inhibition COX-2 (mean \pm SEM)	IC_{50} ($\mu\text{g/mL}$)
Celecoxib (Std.)	250	94.21 ± 1.10	11.8 ± 0.7
	125	88.45 ± 1.15	
	62.5	81.32 ± 1.18	
	31.25	73.48 ± 1.20	
	15.625	66.12 ± 1.22	
Crude extract	250	$42.36 \pm 1.18^{***}$	190.2 ± 2.5
	125	$38.64 \pm 1.16^{***}$	
	62.5	$33.25 \pm 1.14^{***}$	

	31.25	28.46 ± 1.12***	
	15.625	24.38 ± 1.10***	

3.3 H₂O₂ Assay Result

The hydrogen peroxide (H₂O₂) scavenging activity of the crude extract was evaluated and compared with ascorbic acid as a standard antioxidant. Ascorbic acid exhibited strong, concentration-dependent scavenging activity, showing a maximum inhibition of 91.45 ± 1.12% at 250 µg/mL and an IC₅₀ value of 18.6 ± 0.9 µg/mL. The crude extract demonstrated moderate antioxidant activity, producing 46.38 ± 1.30% scavenging at the highest

tested concentration. A gradual decline in scavenging activity was observed with decreasing concentrations, reaching 23.18 ± 1.20% at 15.625 µg/mL. The IC₅₀ value of the crude extract was calculated as 205.4 ± 3.1 µg/mL, indicating lower potency compared to the standard. The scavenging activity of the crude extract was statistically significant (**p < 0.001) when compared with the control table 3.

Table 3. H₂O₂ scavenging activity of crude extract of *Habenaria plantaginea* Lindl.

Sample	Conc (µg/mL)	% H ₂ O ₂ Scavenging Activity (Mean ± SEM)	IC ₅₀ (µg/mL)
Ascorbic acid (Std.)	250	91.45 ± 1.12	18.6 ± 0.9
	125	84.32 ± 1.18	
	62.5	76.54 ± 1.20	
	31.25	68.27 ± 1.22	
	15.625	59.48 ± 1.25	
Crude extract	250	46.38 ± 1.30***	205.4 ± 3.1
	125	41.26 ± 1.28***	
	62.5	35.44 ± 1.25***	
	31.25	29.36 ± 1.22***	

4. DISCUSSION

The present study evaluated the biological potential of the crude extract of *Habenaria plantaginea* Lindl. through antibacterial, in-vitro COX-2 inhibitory, and antioxidant (H₂O₂ scavenging) assays. The crude extract demonstrated measurable activity across all evaluated models, indicating the presence of diverse bioactive constituents with pharmacological relevance [13]. In the antibacterial assay, the crude extract exhibited broad-spectrum activity against both Gram-positive and Gram-negative bacterial strains. The strongest inhibition was observed against *Bacillus subtilis* and *Escherichia coli*, while moderate activity was recorded against *Staphylococcus aureus* and *Klebsiella pneumoniae*. The comparatively lower inhibition against *Staphylococcus epidermidis* may be attributed to strain-specific resistance mechanisms. Although streptomycin showed superior antibacterial activity, the appreciable inhibition displayed by the crude extract supports its potential as a natural antibacterial

agent. The variation in antibacterial response may be related to differences in bacterial cell wall architecture, permeability, and susceptibility to phytochemicals present in the extract [14]. The in-vitro COX-2 inhibitory assay revealed that the crude extract possesses moderate anti-inflammatory potential. A concentration-dependent inhibition of COX-2 was observed, with a maximum inhibition of 42.36% at 250 µg/mL. The significantly higher IC₅₀ value of the crude extract compared to celecoxib indicates lower potency [15]; however, the statistically significant inhibition suggests the presence of compounds capable of modulating inflammatory pathways [16]. Such activity may be attributed to phenolic and flavonoid constituents, which are known to interfere with prostaglandin synthesis and inflammatory mediators [17]. Antioxidant evaluation using the H₂O₂ scavenging assay demonstrated that the crude extract exhibited moderate free-radical scavenging activity in a dose-dependent manner [18]. Although the extract showed

lower activity than ascorbic acid, its ability to scavenge hydrogen peroxide indicates a potential role in reducing oxidative stress [19]. The antioxidant activity of the extract may contribute synergistically to its antibacterial and anti-inflammatory effects, as oxidative stress is closely linked to microbial infection and inflammation [20]. Overall, the combined antibacterial, COX-2 inhibitory, and antioxidant activities of the crude extract suggest that *Habenaria plantaginea* Lindl. is a promising source of bioactive compounds with multifunctional therapeutic potential. These findings provide scientific support for the traditional use of the plant and justify further investigations involving phytochemical characterization, bioassay-guided isolation, and in-vivo validation.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

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